

Short communication

## FISH characterization of t(8;12)(q12;p13) observed as the sole karyotypic anomaly in a myelodysplastic syndrome patient

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### Abstract

We report a t(8;12)(q12; p13) as the sole cytogenetic anomaly in a patient with a myelodysplastic syndrome (MDS). By means of FISH, we mapped the genomic region involved in the breakpoint (bcp) on both chromosomes. The 12p13 bcp mapped between markers WI-664 and WI-9218, immediately distal to the breakpoint cluster region frequently involved in hematological neoplasms targeted by y964C10. The 8q12 bcp (not yet investigated by FISH) was characterized and found to occur between markers WI-3263 and D8S524 within the region recognized by y874E10. © 2001 Elsevier Science Inc. All rights reserved.

### 1. Introduction

Numerical and structural rearrangements of chromosomes 8 and 12 have been described in hematological malignancies as different as acute lymphoblastic (ALL), acute myeloid leukemia (AML), myelodysplastic syndrome (MDS) or myeloproliferative disorders, and non-Hodgkin lymphomas [1]. The breakpoints of translocations and dicentrics involving 12p are most frequently localized in 12p13 within the *ETV6* gene (also known as *TEL*), a member of the ETS family of transcription factors, whose involvement in leukemia is particularly interesting insofar as different translocations are associated with distinct forms of disease [2]. To mention just a few of the partner bands and genes that are affected in different hematological conditions, *ETV6* is fused to the platelet-derived growth factor  $\beta$  receptor in a case of chronic myelomonocytic leukemia carrying a t(5;12) [3] and to the runt-related gene *CBFA2/AML1*, which is affected by the most common t(12;21) in pediatric acute lymphoblastic leukemia [4]. The loss of 12p material is often associated with an *ETV6* translocation and is sometimes accompanied by the concomitant deletion of the other *ETV6* allele [5], a feature linking this rearrangement to a

12p interstitial deletion [6,7]. As far as chromosome 8 is concerned, trisomy 8 is the most frequent numerical aberration in myeloid disorders, including acute myelogenous leukemia (AML), myeloproliferative disorders, and MDS [1,8]. Chromosome translocations are rare in MDS, with the exception of t(8;21)(q12;q22), which is usually found in rapidly evolving MDS [9]. In particular, t(8;12)(q12;p13) has been described in the context of a complex karyotype in a few cases of AML, MDS [6,7,10], and chronic myelogenous leukemia (CML) [6]. We here report the fluorescence in situ hybridization (FISH) characterization of an apparently balanced (8;12)(q12;p13) translocation, identified as the sole cytogenetic lesion in the bone marrow karyotype of a patient with MDS (FAB subtype: refractory anemia with an excess of blasts, RAEB). Using YAC probes located to 8q12 and 12p13, we precisely defined the genomic region involved in the breakpoints on both chromosomes.

### 2. Case report

A 65-year-old female was admitted to our institution in 1994 for pancytopenia. The laboratory data showed anemia (8 g/dl), thrombocytopenia ( $60 \times 10^9/l$ ), and leukopenia ( $2.8 \times 10^9/l$ , with neutrophils  $1.1 \times 10^9/l$ ). A bone marrow aspirate and biopsy detected 9% of myeloid blasts. A diagnosis of RAEB was made on the basis of the FAB Cooperative Group diagnostic criteria. The patient presented with splenomegaly (ultra-

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sound bipolar axis of 20 cm), and was dependent on packed red blood cell transfusions. One course of polychemotherapy with daunorubicin, cytosine arabinoside, and 6-thioguanine induced a complete remission after prolonged bone marrow aplasia complicated by life-threatening pneumonia. In 1995, a splenectomy was performed in an attempt to reduce red blood cell transfusions, but without a lasting response. Between 1995 and 1999, the patient was affected by numerous viral (*herpes zoster*), mycotic (*aspergillus*), and bacterial infections (*pseudomonas*); she died of fungal sepsis in 1999.

### 3. Materials and methods

#### 3.1. Cytogenetics and FISH analysis

Chromosomes were prepared from bone marrow cultures. The cytogenetic analysis was performed using the QFQ banding technique, following the International System for Human Cytogenetic Nomenclature [11].

The YAC probes, used in FISH experiments from the CEPH YAC library [12], were provided by the YAC Screening Centre (Dibit, HSR, Milan, Italy). FISH experiments were performed following the protocols of Lichter et al. [13] and Lichter and Cremer [14], with minor modifications.

### 4. Results

Cytogenetic analysis, performed according to standard methods on 50 metaphases from bone marrow cells, revealed a 46,XX,t(8;12)(q12;p13) acquired karyotype. In order to refine the localization of the 8q12 and 12p13 translocation breakpoints, we performed FISH analyses using yeast artificial chromosome clones (YACs) mapping to these chromosomal bands. Table 1 shows the YAC clones used and the FISH results. The breakpoint on chromosome 8 was investigated by means of 14 overlapping YAC clones belonging to the WC8.7 contig and encompassing the 8q11.2–8q12 bands. The FISH signals of y874E10 were observed on both derivative chromosomes, thus indicating that this YAC encompasses the breakpoint region on the der(8) (Fig. 1a). In order to refine the 8q12 bkp mapping further, we identified two additional YAC clones overlapping the centromeric (y820C4) and telomeric (y849B4) portions of y874E10 (Fig. 1d and Table 1). As y820C4 hybridized only to the der(8) and y849B4 hybridized only to the der(12), we were able to locate the breakpoint between markers WI-3263 and D8S524, which are, respectively, reported to map to the telomeric end of y820C4 and the centromeric end of y849B4 (Table 1, Fig. 1d, and data not shown). To map the chromosome 12 breakpoint, we used six YAC clones anchored to the contig WC12.1 and mapping to 12p13 (Table 1). As FISH signals of y746A12 were detected on the der(8) (Figs. 1b and 1e) and those of y964C10 were detected on the der(12) (Figs. 1c and 1e), the 12p13 breakpoint appears to be delimited by marker WI-9218 mapping to the centromeric end of y746A12 and marker WI-664 mapping to the telomeric end of y964C10 (Fig. 1e). These results were

Table 1  
YAC FISH results on derivative chromosomes

| YAC name            | Localization | FISH signal on derivative chromosomes |
|---------------------|--------------|---------------------------------------|
| 935D11              | 8q11.2       | der(8)                                |
| 787A4               | 8q11.2       | der(8)                                |
| 791E12              | 8q11.2       | der(8)                                |
| 820C4 <sup>a</sup>  | 8q11.2       | der(8)                                |
| 874E10 <sup>a</sup> | 8q12         | der(8)/der(12)                        |
| 849B4 <sup>a</sup>  | 8q12         | der(12)                               |
| 834A9               | 8q12         | der(12)                               |
| 935E9               | 8q12         | der(12)                               |
| 946B7               | 8q12         | der(12)                               |
| 925G7               | 8q12         | der(12)                               |
| 817B1               | 8q12         | der(12)                               |
| 799C11              | 8q12         | der(12)                               |
| 787A6               | 8q12         | der(12)                               |
| 953G3               | 8q12         | der(12)                               |
| 929E11              | 12p13        | der(12)                               |
| 964a12 <sup>a</sup> | 12p13        | der(12)                               |
| 817H1 <sup>a</sup>  | 12p13        | der(12)                               |
| 746A12 <sup>a</sup> | 12p13        | der(8)                                |
| 808E9               | 12p13        | der(8)                                |
| 886D1               | 12p13        | der(8)                                |

YAC order on 8q and 12p is given from tel to cen.

<sup>a</sup>YACs shown in Figs. 1d–1e.

confirmed by hybridization with the contiguous y817H1, which almost completely overlaps y964C10 [15] (Fig. 1e).

### 5. Discussion

A few cases of AML, MDS [6,7,10] and CML [6] have been described harboring the translocation t(8;12)(q12;p13), usually associated with a complex karyotype. We here report a case of MDS (FAB subtype: refractory anemia with an excess of blasts, RAEB), in which a balanced t(8;12)(q12;p13) was the sole cytogenetic lesion in the acquired bone marrow karyotype. Given the possible pathogenetic relevance of unique reciprocal translocations in human leukemia, the regions involved in the breakage, giving rise to the recombinant der(8) and der(12) chromosomes, were characterized by FISH analysis using YAC probes targeting 8q12 and 12p13. This made it possible to map the breakpoints on both partner chromosomes precisely. We identified a molecular probe y874E10 (encompassing the 8q12 breakpoint), which mapped between markers WI-3263 and D8S524. YAC 874E10 may prove to be a useful reagent for FISH characterization of the 8q12 region, which has not yet been targeted in myelodysplasia and AML. We also pinpointed the 12p13 breakpoint between markers WI-664 and WI-9218, which delimit a genomic region immediately telomeric to that identified by y964C10 as containing the *ETV-6* gene that is most frequently involved in deletions and translocations in leukemia [3,6,16]. Two other breakpoint cluster regions, occurring approximately 4 cM distal to the bkp here described, have been reported, thus indicating that the overall 12p13 cytogenetic band is prone to rearrangement [17]. Although y964C10 did not seem to be affected in the translocation of our case, the

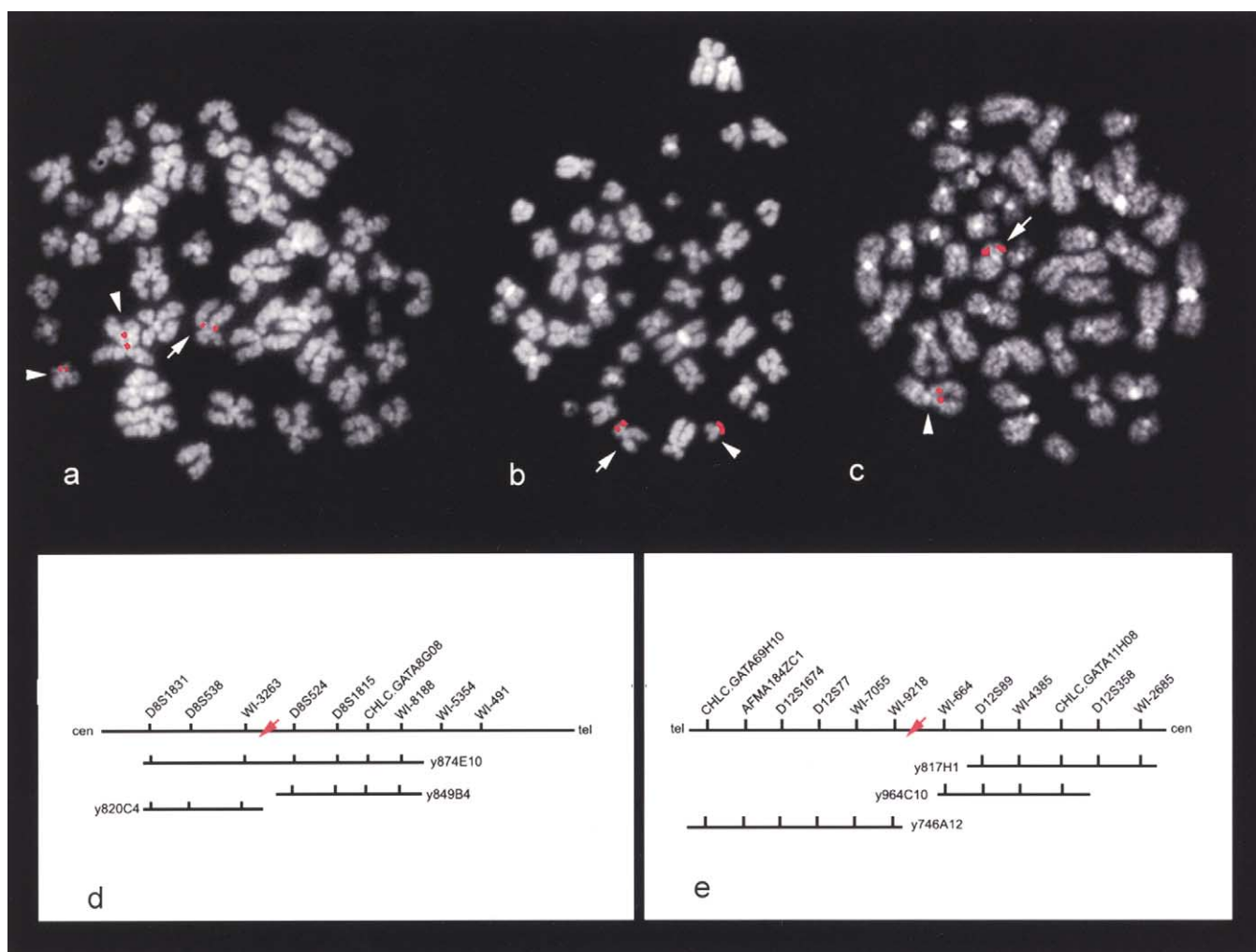


Fig. 1. Characterization by YAC FISH of reciprocal translocation breakpoints leading to der(8) and der(12) chromosomes (a) y874E10 (8q12) shows a split signal on der(8) and der(12) (arrowheads), thus indicating that it spans the bkp on chromosome 8 (normal chromosome 8 identified by arrow); (b) y746A12 mapping to 12p13 shows a "moved" hybridization signal on chromosome der(8) (arrowhead) targeting a region distal to the translocation bkp on 12p13 (normal chromosome 12 indicated by arrow); (c) y964C10, contiguous to y746A12 on 12p13 gives the expected signal on der(12) (arrowhead), thus fixing the centromeric boundary of the translocation bkp on chromosome 12 (arrow). (d) Physical map of the breakpoint regions at 8q12 and (e) at 12p13. The loci are reciprocally ordered as shown in the MIT web sit (<http://www.genome.wi.mit.edu>), but the distances are not to scale. Red arrows point to the regions involved in the breakpoints.

possible involvement of small ETV-6 deletions that escape detection by YAC FISH cannot be ruled out.

In conclusion, our data confirm the association between MDS and 12p13 genetic lesions, particularly t(8;12)(q12;p13). They also provide the first FISH characterization of the 8q12 breakpoint using an 8q12-specific probe that may prove valuable in estimating the frequency of the involvement of this genomic region in MDS. The availability of patients bearing 8q12 rearrangements may allow the refined mapping of the breakpoint and favor the search for candidate genes affected in myelodysplasia.

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#### References

- [1] Mitelman F. Catalog of chromosome aberrations in cancer. New York: Wiley-Liss, 1998.
- [2] Golub TR, Baker GF, Stegmaier K, Gilliland DG. The TEL gene contributes to the pathogenesis of myeloid and lymphoid leukemias by diverse molecular genetic mechanisms. *Curr Top Microbiol Immunol* 1997;220:67–79.
- [3] Golub TR, Baker GF, Lovett M, Gilliland DG. Fusion of PDGF receptor to a novel ets-like gene, tel, in chronic myelomonocytic leukemia with t(5;12) chromosomal translocation. *Cell* 1994;77:307–16.
- [4] Romana SP, Mauchauffe, M, Le Coniat M, Chumakov I, Le Paslier

- D, Berger R, Bernard OA. The t(12;21) of acute lymphoblastic leukemia results in a tel-AML1 gene fusion. *Blood* 1995;85:3662–70.
- [5] Stegmaier K, Reynolds C, Sklar J, Donnelly M, Bohlander SK, Rowley JD, Sallan SE, Gilliland DG, Golub TR. Frequent loss of heterozygosity at the TEL gene locus in acute lymphoblastic leukemia of childhood. *Blood* 1995;86:38–44.
- [6] Sato Y, Bohlander SK, Kobayashi H, Reshmi S, Suto Y, Davis EM, Espinosa III R, Hoopes R, Montgomery KT, Kucherlapati RS, Le Beau MM, Rowley JD. Heterogeneity in the breakpoints in balanced rearrangements involving band 12p13 in hematologic malignancies identified by fluorescence in situ hybridization: TEL (ETV6) is involved in only one half. *Blood* 1997;90:4886–93.
- [7] Streubel B, Sauerland C, Heil G, Freund M, Bartels H, Lengfelder E, Wandt H, Ludwig WD, Nowotny H, Baldus M, Grothaus-Pinke B, Buchner T, Fonatsch C. Correlation of cytogenetic, molecular cytogenetic and clinical findings in 59 patients with ANLL or MDS and abnormalities of the short arm of chromosome 12. *Br J Haematol* 1998;100:521–33.
- [8] Cuneo A, Bigoni R, Roberti MG, Bardi A, Rigolini GM, Piva N, Mancini M, Nanni M, Alimena G, Mecucci C, Matteucci C, La Starza R, Bernasconi P, Cavigliano P, Genini E, Zaccaria A, Testoni N, Carboni C, Castoldi G. Detection and monitoring of trisomy 8 by fluorescence in situ hybridization in acute myeloid leukemia: a multicentric study. *Haematologica* 1998;83:21–6.
- [9] Mecucci C, La Starza R. Cytogenetics of myelodysplastic syndromes. *Forum* 1999;9:4–13.
- [10] Hernandez JM, Gonzalez MB, Garcia JL, Ferro MT, Gutierrez NC, Marynen P, San Miguel JF. Two cases of myeloid disorders and a t(8;12)(q12;p13). *Haematologica* 2000;85:31–4.
- [11] ISCN. An international system for human cytogenetic nomenclature. F Mitelman, editor. Basel: S. Karger, 1995.
- [12] Chumakov IM, Rigault P, Le Gall I, Bellane-Chantelot C, Cohen D. A YAC contig map of the human genome. *Nature* 1995;377(Suppl 28):175–297.
- [13] Lichter P, Tang Chang C-J, Call K, Hermanson G, Evans GA, Housman D, Ward C. High resolution mapping of human chromosome 11 by in situ hybridization with cosmid clones. *Science* 1990;247:64–9.
- [14] Lichter P, Cremer T. Chromosome analysis by non-isotopic in situ hybridization. In: Rooney DE, Czepulkowski BH, editors. *Human cytogenetics—a practical approach*. New York: Oxford University Press, 1992, pp. 157–92.
- [15] Baens M, Wlodarska I, Corveleyn A, Hoornaert I, Hagemijer A, Marynen P. A physical, transcript and deletion map of chromosome region 12p12.3 flanked by ETV6 and CDKN1B: hypermethylation of the LRP6 CpG island in two leukemia patients with hemizygous del(12p). *Genomics* 1999;56:40–50.
- [16] Kobayashi H, Montgomery KT, Bohlander SK, Adra CN, Lim BL, Kucherlapati RS, Donis-Keller H, Holt MS, Le Beau MM, Rowley JD. Fluorescence in situ hybridisation mapping of translocations and deletions involving the short arm of human chromosome 12 in malignant hematological malignancies. *Blood* 1994;84:3473–9.
- [17] Kucherlapati R, Marynen P, Turc-Carel C. Report of the Fourth International Workshop on Human Chromosome 12 Mapping. *Cytogenet Cell Genet* 1997;78:82–95.